Noninvasive Prenatal Aneuploidy Screen by Cell-Free DNA Sequencing

Cell-free DNA (cfDNA) screening, also referred to as noninvasive prenatal testing (NIPT) or noninvasive prenatal screening (NIPS), is the most sensitive and specific prenatal screening option for trisomy 13, trisomy 18, trisomy 21 (Down syndrome), and Turner syndrome. The American College of Obstetricians and Gynecologists (ACOG), American College of Medical Genetics and Genomics (ACMG), and Society for Maternal-Fetal Medicine (SMFM) recommend that prenatal genetic screening, including NIPT, and prenatal diagnosis be offered to all pregnant women regardless of maternal age or fetal risk of chromosomal abnormality. High-risk results merit prompt, appropriate follow-up; critical clinical decisions should be based on diagnostic rather than screening test results. Guidelines recommend discussing the option of diagnostic testing (amniocentesis or chorionic villus sampling) instead of screening and after high-risk screening results. This is particularly important for women whose clinical and/or family history places them at high risk for genetic conditions not targeted by NIPT. For more details about NIPT and a description of the spectrum of prenatal genetic testing, please refer to the ARUP Consult Prenatal Testing for Chromosomal Abnormalities and Neural Tube Defects topic. For more information about maternal serum screening, refer to ARUP's First and Second Trimester Screening Options.

Disease Overview

Aneuploidy is typically due to nondisjunction (uneven separation) of chromosome pairs/copies during gametogenesis (sperm and egg formation by meiosis). This leads to one or three copies of a chromosome in the zygote (fertilized egg), which is then replicated by mitosis into each cell of the body, causing disruption of the normal course of development and cellular function. The prevalence of aneuploidy is higher in early gestation pregnancies than in term pregnancies. The table below details specific disorders and their associated clinical findings and incidences.

<table>
<thead>
<tr>
<th>Disorder</th>
<th>Clinical Features</th>
<th>Incidence at Birth</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trisomy 13 (Patau syndrome)</td>
<td>Clinical features include multiple congenital malformations and severe to profound intellectual disability&lt;br&gt;Babies with trisomy 13 have high prenatal, neonatal, and infant mortality; approximately 10% survive past 5 years of age</td>
<td>1/5,000</td>
</tr>
<tr>
<td>Trisomy 18 (Edwards syndrome)</td>
<td>Clinical features include intrauterine growth restriction with low birth weight; multiple congenital anomalies involving the brain, spinal cord, heart, abdominal wall, and kidneys; hypotonia; and severe to profound intellectual disability&lt;br&gt;Babies with trisomy 18 have high prenatal, neonatal, and infant mortality; approximately 12% survive past 5 years of age</td>
<td>1/3,330</td>
</tr>
<tr>
<td>Trisomy 21 (Down syndrome)</td>
<td>Clinical features include hypotonia, a characteristic facial appearance, developmental delays/intellectual disability, and short stature</td>
<td>1/660</td>
</tr>
<tr>
<td>Monosomy X (Turner syndrome)</td>
<td>Clinical features include fluid accumulations (cystic hygroma, lymphedema), short stature, low-set ears, broad chest, and cardiac and renal anomalies&lt;br&gt;Infertility is typical&lt;br&gt;Intelligence is generally within the normal range</td>
<td>1/2,500 female births</td>
</tr>
<tr>
<td>XXY syndrome (Klinefelter syndrome)</td>
<td>Males with Klinefelter syndrome, or 47,XXX, are typically taller than average, with delayed or incomplete puberty and infertility&lt;br&gt;The effect on social skills and learning is widely variable and often mild</td>
<td>1/500 male births</td>
</tr>
</tbody>
</table>

Sources: Jones, 2013; Meyer, 2016; Otter, 2010; Unique; Bardsley, 2013; Groth, 2013
**Disorder** | **Clinical Features** | **Incidence at Birth**
--- | --- | ---
XYY syndrome | Males with XYY syndrome and females with triple X syndrome (trisomy X) have tall stature and normal physical development. They are fertile and cognition is typically within normal range. | 1/800 male births
XXX syndrome | They have some susceptibility to learning disabilities and other neurocognitive difficulties. | 1/1,000 female births

Sources: Jones, 2013; Meyer, 2016; Otter, 2010; Unique; Bardsley, 2013; Groth, 2013

**Inheritance and Recurrence Risk**

Most cases of trisomy and Turner syndrome are not inherited. Even in sporadic cases, however, the recurrence risk is usually increased over the background risk. Therefore, offering prenatal diagnosis in future pregnancies is warranted. 12,13

**Test Description**

NIPT uses massively parallel whole genome sequencing of cfDNA fragments derived from maternal peripheral whole blood samples to generate paired-end sequencing data that are aligned to a reference genome. Regions of the genome that are over- or underrepresented are quantified. The probability (log likelihood ratio) of aneuploidy is calculated for targeted chromosomes, accounting for bias of GC-rich DNA fragments, region-specific genomic coverage, and fetal fraction. The results are reported as “high risk” or “low risk” for the aneuploidies of interest. 14,15,16

Fetal fraction is estimated from the distribution of cfDNA fragment lengths and their genomic coordinates. A dynamic assessment of data quality and coverage profile determines whether data quality is sufficient for aneuploid risk interpretation.14 Refer to the ARUP NIPT patient brochure for more information.

ARUP prohibits the use of Non-Invasive Prenatal Aneuploidy Screen by Cell-Free DNA Sequencing to further discrimination based on genetic characteristics, oppression of at-risk or minority populations, clinical utilization of human heritable genome editing, reproductive cloning of human beings, and/or bioterrorism.

**Test Interpretation**

**Published Clinical Sensitivity and Specificity of NIPT**

<table>
<thead>
<tr>
<th>Condition</th>
<th>Clinical Sensitivity</th>
<th>Clinical Specificity</th>
<th>PPVC</th>
<th>NPV</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trisomy 21</td>
<td>98.9%</td>
<td>99.7%</td>
<td>20.9-95.1%</td>
<td>&gt;99.9%</td>
</tr>
<tr>
<td>Trisomy 18</td>
<td>&gt;99.9%</td>
<td>99.8%</td>
<td>6.2-94.1%</td>
<td>&gt;99.9%</td>
</tr>
<tr>
<td>Trisomy 13</td>
<td>&gt;99.9%</td>
<td>99.8%</td>
<td>1.6%-76.8%</td>
<td>&gt;99.9%</td>
</tr>
<tr>
<td>Monosomy X</td>
<td>&gt;99.9%</td>
<td>&gt;99.8%</td>
<td>—</td>
<td>—</td>
</tr>
</tbody>
</table>

a Male/female concordance is >99.
b Data are not sufficient to calculate sensitivity, specificity, PPV, and NPV for every condition.
c PPV ranges are calculated based on sensitivity and specificity in Borth17 and the prevalence in a low-risk group (20 years of age, 30 weeks gestational age) through the prevalence in a high-risk group (44 years of age, 10 weeks gestational age). PPV is greatly affected by an individual’s pretest risk for each of the screened conditions, which varies based on gestational age and maternal age.

d NPV, negative predictive value; PPV, positive predictive value
Sources: Borth, 2021; Snijders, 1991; Snijders, 1995

Detailed information about PPV at increased pretest risk levels, various gestational ages, and maternal ages can be found in the Positive Predictive Value of a High-Risk Noninvasive Prenatal Screening Result for Various Increased Pretest Risk Levels and for Various Gestational Ages and Maternal Ages tables.
# Analytic Validation Accuracy

## Positive Percent Agreement (Sensitivity) and Negative Percent Agreement (Specificity)

<table>
<thead>
<tr>
<th>Condition or Characteristic</th>
<th>Mean PPA (Sensitivity)</th>
<th>5% to 95% Confidence Interval</th>
<th>Number of Samples</th>
<th>PPV</th>
<th>NPA (Specificity)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trisomy 21</td>
<td>94.9%</td>
<td>82.6%-99.3%</td>
<td>59</td>
<td>38.6%-97.5%</td>
<td>89.7%-99.9%</td>
</tr>
<tr>
<td>Trisomy 18</td>
<td>94.8%</td>
<td>78.6%-99.6%</td>
<td>39</td>
<td>8.6%-95.8%</td>
<td>94.8%-99.9%</td>
</tr>
<tr>
<td>Trisomy 13</td>
<td>86.7%</td>
<td>52.3%-98.9%</td>
<td>15</td>
<td>2.7%-85.0%</td>
<td>98.2%-99.9%</td>
</tr>
<tr>
<td>Monosomy X</td>
<td>&gt;99.9%</td>
<td>64.1%-100%</td>
<td>10</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>XXX</td>
<td>&gt;99.9%</td>
<td>34.4%-100%</td>
<td>4</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>XXY</td>
<td>80.0%</td>
<td>36.7%-98.4%</td>
<td>10</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>XYY</td>
<td>85.7%</td>
<td>34.5%-99.7%</td>
<td>7</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Male</td>
<td>&gt;99.9%</td>
<td>97.29%-100%</td>
<td>166</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Female</td>
<td>99.3%</td>
<td>95.3%-99.9%</td>
<td>149</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td><strong>PPA (Sensitivity)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Combined aneuploidy</td>
<td>93.1%</td>
<td>85.5%-97.4%</td>
<td>144</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td><strong>NPA (Specificity)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Euploid/low risk</td>
<td>99.5%</td>
<td>96.2%-99.9%</td>
<td>186</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

*PPA and NPA are commonly referred to as sensitivity and specificity; in this validation the terms PPA and NPA are more appropriate.

*For samples with observed fetal fraction of ≤5.0%, the sensitivity to detect fetal aneuploidy and accuracy of the fetal fraction estimate are significantly lower.

*Data are not sufficient to calculate PPV and NPA for every condition.

*Jeffreys interval.

*PPV is greatly affected by an individual's pretest risk for each of the screened conditions, which varies based on gestational age and maternal age.

NPA, negative percent agreement; PPA, positive percent agreement

Sources: Snidjers, 1999; Snidjers, 1995

## Reporting Information

- **Autosomal aneuploidies**
  - The following autosomal aneuploidies are reported:
    - Trisomy 13
    - Trisomy 18
    - Down syndrome (trisomy 21)
- **Sex chromosome anomalies (SCA)**
  - The following SCAs are reported in singleton pregnancies:
    - XO (Turner syndrome)
    - XXY (Klinefelter syndrome)
    - XXX
    - XYY
- **Fetal sex**
  - Fetal sex is reported; opting out of fetal sex reporting is available.
- **Fetal fraction estimation (FFE)**
  - FFE is reported as a percentage (standard deviation, 1.2%).
- **Risk results**
The accuracy, or PPV, of NIPT varies depending on the pretest risk for the assessed condition. For more information, please refer to Positive Predictive Value (PPV) of a High-Risk Noninvasive Prenatal Screening Result for Various Increased Pretest Risk Levels and for Various Gestational Ages and Maternal Ages.

An over- or underrepresentation of targeted chromosomes is reported as a high-risk or low-risk result.

High risk for autosomal aneuploidy reports include a PPV statement.

High risk for SCA reports include data from a large, prospective study to aid in counseling patients. If the result is high risk for fetal SCA, this interpretation is reported even if the fetal sex reporting selection is "No." Sex of the fetus may be inferred.

Low-risk reports include negative likelihood ratios for trisomies 13, 18, and 21, calculated by Gil, 2017.

Follow-up recommendations are provided for high-risk results and cases of "no-call" results.

Enhanced reports

Enhanced reports for high-risk autosomal aneuploidy results include a PPV table.

Limitations

- Aneuploidy for higher-order multiple gestations (triplets) is not assessed.
- Fetal demise/miscarriage is not assessed.
- NIPT is a screening test; it is not a diagnostic test. Therefore, results should not be the sole basis for any pregnancy management decision.
- Results should be considered in the context of other clinical findings and test results.
- A low-risk screening result does not exclude a fetal diagnosis of aneuploidy.
- A high-risk screening result does not confirm a fetal diagnosis of aneuploidy.
- Analysis may fail due to a combination of insufficient fetal fraction and dynamic quality metrics including data quality and coverage profile.
- A result may be unreportable due to findings that are uninterpretable or outside the scope of the test.
- Only the risk for fetal aneuploidy involving chromosomes 13, 18, 21, X, and Y is assessed.
- The VeriSeq NIPT Solution v2 is not intended to detect the following conditions:
  - Aneuploidies involving chromosomes not interrogated by this test
  - Polyploidy, such as triploidy
  - Deletions or duplications within any chromosome
  - Balanced chromosome rearrangements
  - Copy number variants of any chromosome
  - Other genetic disorders
  - Birth defects (e.g., open neural tube defect)
- NIPT analysis may yield unexpected results that indicate abnormalities in maternal cfDNA and/or other fetoplacental conditions, such as mosaicism for the targeted chromosomes.
- The results of the test can be confounded by certain maternal and fetal factors including, but not limited to, the following:
  - Recent maternal blood transfusion
  - Maternal organ transplant
  - Maternal surgical procedure
  - Maternal immunotherapy or stem cell therapy
  - Maternal malignancy
  - Maternal aneuploidy or mosaicism
  - Fetoplacental mosaicism
  - Fetal demise or nonviable twin

References


Related Information

Maternal Serum Screening
Prenatal Testing for Chromosomal Abnormalities and Neural Tube Defects

Related Tests

Maternal Serum Screen, Alpha Fetoprotein 3000144
Method: Quantitative Chemiluminescent Immunoassay

Chromosome Analysis, Amniotic Fluid 2002293
Method: Giemsa Band

Chromosome Analysis, Chorionic Villus 2002291
Method: Giemsa Band

Chromosome FISH, Amniotic Fluid with Reflex to Chromosome Analysis or Genomic Microarray 2011130
Method: Fluorescence in situ Hybridization (FISH)

Chromosome FISH, Chorionic Villus with Reflex to Chromosome Analysis or Genomic Microarray 2011131
Method: Fluorescence in situ Hybridization (FISH)

Chromosome Analysis, Amniotic Fluid, with Reflex to Genomic Microarray 2008367
Method: Giemsa Band/Genomic Microarray (Oligo-SNP Array)

Cytogenomic SNP Microarray - Fetal 2002366
Method: Genomic Microarray (Oligo-SNP Array)