

Client: Example Client ABC123  
123 Test Drive  
Salt Lake City, UT 84108  
UNITED STATES

Physician: Doctor, Example

**Patient: Patient, Example**

**DOB:** 1/25/2004  
**Gender:** Female  
**Patient Identifiers:** 01234567890ABCD, 012345  
**Visit Number (FIN):** 01234567890ABCD  
**Collection Date:** 00/00/0000 00:00

**Primary Antibody Deficiency Panel, Sequencing and Deletion/Duplication**

ARUP test code 2011156

Primary Antibody Deficiency Specimen      whole Blood

Primary Antibody Deficiency Panel Interp

Negative

**RESULT**  
No pathogenic variants were detected in any of the genes tested.

**INTERPRETATION**  
No pathogenic variants were detected in any of the genes tested. This result decreases the likelihood of, but does not exclude, a heritable primary antibody deficiency. Please refer to the background information included in this report for a list of the genes analyzed, methodology, and limitations of this test.

**RECOMMENDATIONS**  
Medical screening and management should rely on clinical findings and family history. If this individual has a family history, determination of a causative familial variant in an affected family member is necessary for optimal interpretation of this negative result. Further testing may be warranted if there is a familial variant that is not detectable by this assay. Genetic consultation is recommended.

**COMMENTS**  
Likely benign and benign variants are not reported. Variants in the following region(s) may not be detected by NGS with sufficient confidence in this sample due to technical limitations: None

This result has been reviewed and approved by [REDACTED]

**BACKGROUND INFORMATION:** Primary Antibody Deficiency Panel, Sequencing and Deletion/Duplication

**CHARACTERISTICS:** Primary antibody deficiencies are a group of genetic disorders affecting antibody production. Three categories of primary antibody deficiencies include common variable immunodeficiency disorders (CVID), agammaglobulinemia, and hyper-IgM syndrome.

**EPIDEMIOLOGY:** Incidence is estimated at 1 in 10,000.

**INHERITANCE:** X-linked, autosomal dominant, or autosomal recessive; dependent on the causative gene

**CLINICAL SENSITIVITY:** Approximately 20 percent for CVID; 75-80 percent for hyper-IgM syndrome; 90 percent for agammaglobulinemia; unknown for other syndromes

**GENES TESTED:** ADA; ADA2; AICDA; ATM; ATP6AP1; BLNK; BTK; CARD11;

**H=High, L=Low, \*=Abnormal, C=Critical**

CD19; CD27; CD40; CD40LG; CD70; CD79A; CD79B; CDCA7; CR2; CTLA4; CXCR4\*; DCLRE1C\*; DNMT3B; GATA2; HELLS; ICOS; IGHM; IGLL1; IKZF1; IL21R; KDM6A; KMT2D; LRBA; MOGS; MS4A1; NBN; NFKB1; NFKB2; NFKBIA\*\*; PIK3CD; PIK3R1; PLCG2; PRKCD\*; RAC2; RAG1; RAG2; RNF168; SH2D1A; STAT3; TCF3\*\*; TNFRSF13B; TRNT1; TTC37; UNG; XIAP\*; ZBTB24

\*One or more exons are not covered by sequencing for the indicated gene; see limitations section below.

\*\*Deletion/duplication analysis is not available for this gene.

**METHODOLOGY:** Probe hybridization-based capture of all coding exons and exon-intron junctions of the targeted genes, followed by massively parallel sequencing. Sanger sequencing was performed as necessary to fill in regions of low coverage and to confirm reported variants that do not meet acceptable quality metrics. A proprietary bioinformatic algorithm was used to detect large (single exon-level or larger) deletions or duplications in the indicated genes. Large deletions/duplications confirmed using an orthogonal exon-level microarray. Human genome build 19 (Hg 19) was used for data analysis.

**ANALYTICAL SENSITIVITY:** The analytical sensitivity is approximately 99 percent for single nucleotide variants (SNVs) and greater than 93 percent for insertions/duplications/deletions (indels) from 1-10 base pairs in size. Indels greater than 10 base pairs may be detected, but the analytical sensitivity may be reduced. Deletions of 2 exons or larger are detected with sensitivity greater than 97 percent; single exon deletions are detected with 62 percent sensitivity. Duplications of 3 exons or larger are detected at greater than 83 percent sensitivity. Specificity is greater than 99.9 percent for all variant classes.

**LIMITATIONS:** A negative result does not exclude a primary antibody deficiency. This test only detects variants within the coding regions and intron-exon boundaries of the targeted genes. Deletions/duplications/insertions of any size may not be detected by massively parallel sequencing. Regulatory region variants and deep intronic variants will not be identified. Precise breakpoints for large deletions or duplications are not determined in this assay and single exon deletions/duplications may not be detected based on the breakpoints of the rearrangement. The actual breakpoints for the deletion or duplication may extend beyond or be within the exon(s) reported. This test is not intended to detect duplications of 2 or fewer exons in size, though these may be identified. Single exon deletions are reported but called at a lower sensitivity. Diagnostic errors can occur due to rare sequence variations. In some cases, variants may not be identified due to technical limitations caused by the presence of pseudogenes, repetitive, or homologous regions. This test is not intended to detect low-level mosaic or somatic variants, gene conversion events, complex inversions, translocations, mitochondrial DNA (mtDNA) mutations, or repeat expansions. Interpretation of this test result may be impacted if this patient has had an allogeneic stem cell transplantation. Noncoding transcripts were not analyzed.

The following regions are not sequenced due to technical limitations of the assay:

CXCR4 (NM\_001348056, NM\_001348059) exon(s) 2  
DCLRE1C (NM\_001350965) exon(s) 15  
DCLRE1C (NM\_001350966) exon(s) 13  
DCLRE1C (NM\_001350967) exon(s) 16  
PRKCD (NM\_001354676, NM\_001354678) exon(s) 1  
XIAP (NM\_001167, NM\_001204401, NM\_001378590, NM\_001378591, NM\_001378592) exon(s) 4

Single exon deletions/duplications will not be called for the following exons:

ADA (NM\_000022, NM\_001322051) 1; CXCR4 (NM\_001348056) 2; CXCR4

**H=High, L=Low, \*=Abnormal, C=Critical**

(NM\_001348059) 2; DCLRE1C (NM\_001033855) 4-9; DCLRE1C (NM\_001033857, NM\_001289077) 6-10; DCLRE1C (NM\_001033858, NM\_001289079) 7-11; DCLRE1C (NM\_001289076, NM\_001289078) 3-7; DCLRE1C (NM\_001350965) 4-9,15; DCLRE1C (NM\_001350966) 3-7,13; DCLRE1C (NM\_001350967) 6-10,16; DCLRE1C (NM\_022487) 4-8; HELLS (NM\_018063, NM\_001289067, NM\_001289068, NM\_001289069, NM\_001289070, NM\_001289072) 7; HELLS (NM\_001289071) 8; HELLS (NM\_001289073) 6; IGLL (NM\_152855) 2; IKZF1 (NM\_001291846, NM\_001291847) 4; MOGS (NM\_001146158) 2; PRKCD (NM\_001354676, NM\_001354678) 1; XIAP (NM\_001167, NM\_001204401, NM\_001378590, NM\_001378592) 4; XIAP (NM\_001378591) 5

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the U.S. Food and Drug Administration. This test was performed in a CLIA-certified laboratory and is intended for clinical purposes.

Counseling and informed consent are recommended for genetic testing. Consent forms are available online.

VERIFIED/REPORTED DATES

Procedure	Accession	Collected	Received	Verified/Reported
Primary Antibody Deficiency Specimen	23-356-400513	00/00/0000 00:00	00/00/0000 00:00	00/00/0000 00:00
Primary Antibody Deficiency Panel Interp	23-356-400513	00/00/0000 00:00	00/00/0000 00:00	00/00/0000 00:00

END OF CHART

**H=High, L=Low, \*=Abnormal, C=Critical**