

Client: Example Client ABC123
123 Test Drive
Salt Lake City, UT 84108
UNITED STATES

Physician: Doctor, Example

Patient: Patient, Example

DOB: Unknown
Gender: Unknown
Patient Identifiers: 01234567890ABCD, 012345
Visit Number (FIN): 01234567890ABCD
Collection Date: 00/00/0000 00:00

Holoprosencephaly Panel, Sequencing and Deletion/Duplication

ARUP test code 2008848

Holoprosencephaly Panel Specimen whole Blood

Holoprosencephaly Panel Interp

Positive

INDICATION FOR TESTING
Multiple congenital anomalies, including alobar holoprosencephaly, cleft lip/palate, and hypotelorism

RESULT
One pathogenic variant was detected in the ZIC2 gene.

PATHOGENIC VARIANT
Gene: ZIC2 (NM_007129.3)
Nucleic Acid Change: c.1277del; heterozygous
Amino Acid Alteration: p.Pro426ArgfsTer129
Inheritance Pattern: Autosomal dominant

INTERPRETATION
One copy of a pathogenic variant, c.1277del; p.Pro426ArgfsTer129, was detected in the ZIC2 gene by massively parallel sequencing. Pathogenic ZIC2 variants are associated with holoprosencephaly 5 (MIM: 609637). This molecular result is consistent with a diagnosis of holoprosencephaly.

Please refer to the background information included in this report for a list of the genes analyzed, methodology, and limitations of this test.

Evidence for variant classification:
The ZIC2 c.1277del; p.Pro426ArgfsTer129 variant has been previously identified in a cohort of holoprosencephaly (HPE) patients (Roessler, 2009), and is reported in an additional female patient described as having semilobar HPE (Solomon, 2010). This variant is absent from the Genome Aggregation Database, indicating it is not a common polymorphism. This variant results in a premature termination codon in the last exon of the ZIC2 gene. While this may not lead to nonsense-mediated decay, it is expected to create a truncated ZIC2, and frameshift variants occurring after this variant have also been identified in HPE patients (Roessler, 2009). Based on available information, this variant is considered to be pathogenic.

RECOMMENDATIONS
Genetic consultation is indicated, including a discussion of medical screening and management. At-risk family members should be offered testing for the identified pathogenic ZIC2 variant (Familial Mutation, Targeted Sequencing, ARUP test code 2001961).

H=High, L=Low, *=Abnormal, C=Critical

COMMENTS

Benign and likely benign variants are not reported. Variants in the following region(s) may not be detected by NGS with sufficient confidence in this sample due to technical limitations: None

REFERENCES

Roessler E, et al. The full spectrum of holoprosencephaly-associated mutations within the ZIC2 gene in humans predicts loss-of-function as the predominant disease mechanism. Hum Mutat. 2009;30(4):E541-54.

Solomon BD et al. Mutations in ZIC2 in human holoprosencephaly: description of a novel ZIC2 specific phenotype and comprehensive analysis of 157 individuals. J Med Genet. 2010 Aug;47(8):513-524.

This result has been reviewed and approved by Laura Livnat
BACKGROUND INFORMATION: Holoprosencephaly Panel

CHARACTERISTICS: Holoprosencephaly (HPE) originates from failed midline delineation during early embryonic development and results in partial or complete failure of the prosencephalon to divide into hemispheres. Subtypes of HPE are based on the degree of brain separation and are typically classified as lobar, semilobar, alobar, and middle interhemispheric variant. Microform HPE is an additional subtype that has HPE-related craniofacial anomalies without structural brain defects. The range of clinical findings varies significantly within and between each subtype, including craniofacial characteristics, developmental delay, and neurological impacts. HPE may be an isolated finding or part of a broader syndrome.

EPIDEMIOLOGY: The incidence is 1 in 10,000 live births.

CAUSE: 25-60 percent of cases are due to underlying cytogenetic abnormalities, including numerical or structural chromosome anomalies and pathogenic copy number variations. Pathogenic germline variants in single genes account for 25 percent of cases.

INHERITANCE: Dependent on etiology; autosomal dominant for genes tested in this panel

PENETRANCE: Incomplete with highly variable expression
CLINICAL SENSITIVITY: 25 percent

GENES TESTED: CDON; FGFR1*; GLI2; PTCH1; SHH; SIX3; TGIF1; ZIC2*
*One or more exons are not covered by sequencing and/or deletion/duplication analysis for the indicated gene; see limitations section below.

METHODOLOGY: Probe hybridization-based capture of all coding exons and exon-intron junctions of the targeted genes, followed by massively parallel sequencing. Sanger sequencing was performed as necessary to fill in regions of low coverage and to confirm reported variants that do not meet acceptable quality metrics. A proprietary bioinformatic algorithm was used to detect large (single exon-level or larger) deletions or duplications in the indicated genes. Large deletions/duplications confirmed using an orthogonal exon-level microarray. Human genome build 19 (Hg 19) was used for data analysis.

ANALYTICAL SENSITIVITY/SPECIFICITY: The analytical sensitivity is approximately 99 percent for single nucleotide variants (SNVs) and greater than 93 percent for insertions/duplications/deletions (indels) from 1-10 base pairs in size. Indels greater than 10 base pairs may be detected, but the analytical sensitivity may be reduced. Deletions of 2 exons or larger are detected with sensitivity greater than 97 percent; single exon deletions are detected with 62 percent sensitivity.

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Duplications of 3 exons or larger are detected at greater than 83 percent sensitivity. Specificity is greater than 99.9 percent for all variant classes.

LIMITATIONS: A negative result does not exclude a heritable form of holoprosencephaly. This test only detects variants within the coding regions and intron-exon boundaries of the targeted genes. Deletions/duplications/insertions of any size may not be detected by massively parallel sequencing. Regulatory region variants and deep intronic variants will not be identified. Precise breakpoints for large deletions or duplications are not determined in this assay and single exon deletions/duplications may not be detected based on the breakpoints of the rearrangement. The actual breakpoints for the deletion or duplication may extend beyond or be within the exon(s) reported. This test is not intended to detect duplications of 2 or fewer exons in size, though these may be identified. Single exon deletions are reported but called at a lower sensitivity. Diagnostic errors can occur due to rare sequence variations. In some cases, variants may not be identified due to technical limitations caused by the presence of pseudogenes, repetitive, or homologous regions. This test is not intended to detect low-level mosaic or somatic variants, gene conversion events, complex inversions, translocations, mitochondrial DNA (mtDNA) mutations, or repeat expansions. Interpretation of this test result may be impacted if this patient has had an allogeneic stem cell transplantation. Noncoding transcripts were not analyzed.

The following regions are not sequenced due to technical limitations of the assay:

FGFR1 (NM_001354367) exon(s) 18
FGFR1 (NM_001354369) exon(s) 18
FGFR1 (NM_001354370) exon(s) 17
ZIC2 (NM_007129) partial exon(s) 3(Chr13:100637736-100637843)
Single exon deletions/duplications will not be called for the following exons: FGFR1 (NM_001354367, NM_001354369) 18; FGFR1 (NM_001354370) 17

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the U.S. Food and Drug Administration. This test was performed in a CLIA-certified laboratory and is intended for clinical purposes.

Counseling and informed consent are recommended for genetic testing. Consent forms are available online.

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VERIFIED/REPORTED DATES

Procedure	Accession	Collected	Received	Verified/Reported
Holoprosencephaly Panel Specimen	24-212-111463	00/00/0000 00:00	00/00/0000 00:00	00/00/0000 00:00
Holoprosencephaly Panel Interp	24-212-111463	00/00/0000 00:00	00/00/0000 00:00	00/00/0000 00:00

END OF CHART

H=High, L=Low, *=Abnormal, C=Critical

Unless otherwise indicated, testing performed at:

ARUP LABORATORIES | 800-522-2787 | aruplab.com
500 Chipeta Way, Salt Lake City, UT 84108-1221
Jonathan R. Genzen, MD, PhD, Laboratory Director

Patient: Patient, Example
ARUP Accession: 24-212-111463
Patient Identifiers: 01234567890ABCD, 012345
Visit Number (FIN): 01234567890ABCD
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